

Please send completed form to the C3:

ATTN: Christine Lafontaine

Email: <u>c3@ohri.ca</u>

Phone:+1(613)798-5555 ext 73860

Fax: +1(613)737-8659



PATIENT INFORMATION					
Name (Last, First)					
Medical Record #					
Date of Birth (YYYY/MM	/DD):	Gender: M F			
Address:		City:			
Province:	Country:	Postal/Zip code			

FdX: +1(013)/3/-8059		Province:	Country:	Postal/Zip code	
	ORDER	INFORMATION			
Requesting Physician Location/Facility: Phone: Secondary contact (office assistant/nurse) Phone	Report	delivery method:	Email Fax		
DIAGNOSIS & ELIGIBILITY CRITERIA					
Biliary Tract Cancer Diagnosis (select all that a lateral Intrahepatic Extrahepatic Extrahepatic Equired Eligibility Criteria: 1. Age 18 or older YES 2. Unresectable locally advanced OR Metast 3. Patient agrees to contact the C3 (c3@ohri	Perihilar Dis	Conditional Eligi 1. Being consider 2. Has already	adder Unconfi ibility Criteria (See page dered for clinical trials? had molecular testing? py of report is required)		
	TES	T REQUEST			
	GENOMIC I	PANEL ORDERING:			
OncoHelix-2 170 genes CGP Panel (Tissue: DNA & RNA		_	l *see page 3 for details DNA); <u>Fusions:</u> 55 RNA fu	sion genes; <u>CNV:</u> 59 targets	
	SPECIM	EN RETRIEVAL			
OncoHelix Navigator will contact Pathology Lab		Copy of pathology	to be provided by Order report attached:	YES (required)	
Pathologist Name:	Pathology Lab:	Phone:		Fax:	
Specimen ID:	Specimen Site:	С	Date of Collection (YYYY/I	MM/DD):	
	TEST AUTHORIZATIO	N, CONSENT & SIGNAT	URES		
I certify that I am the patient's treating physicial purpose of testing to the patient and have obtained retain test results indefinitely for internal quality ass and sequencing data for ongoing/future unspecified	d informed consent, to the ex- surance/operational improver research and development p	ttent legally required, to p ment, and (c) use/disclose urposes.	permit OncoHelix to (a) perf de-identified (without iden	orm the test/s specified herein, (b) tifiable patient information) results	
Ordering Physician signature	Printed Name		Da	te	
I permit OncoHelix partner lab HTL to (a) perform the test/s specified herein, that may include de-identified sequencing data analysis in the US and Europe with final analysis in Canada (b) retain de-identified test results as required or permitted by law for internal quality assurance / operational improvement, (c) use/disclose de-identified results and sequencing data for ongoing/future unspecified research and development purposes, (d) share de-identified aggregate data to the funder of the program C3/OHRI ("Funder") for use in reporting, submissions, publication, research or commercial purposes or to improve this program. The Funder may modify or terminate the program at any time in its sole discretion. I acknowledge that I need to contact the C3 before testing can be completed. Should I not reach out in a timely manner, I permit the C3 to contact me.					
Patient's signature OR Patient Verbal Consent Obtained from Orde	Printed Name	Conta	act information r phone number)	Date	
	3				





SAMPLE REQUIREMENT & GUIDELINES

Nucleic Acid and Tissue for Solid Tumor Genomic Analysis Panels						
Panel	DNA	RNA	Biopsy	FFPE		Guidelines for 170 panel
OncoHelix-2 170 genes CGP Panel	250 ng	150 ng	120 μm or 4 mm ³	✓	embedded (FFPE) t • 120 μm of FFPE t 40% tissue content	c acids and fresh frozen (FF) or formalin fixed paraffin issue samples are accepted cissue section (4 scrolls of 30 µm thickness) with a minimum of ta 10% tumor cellularity; or 2-4 FFPE cores of 1-2 mm³; or 4 DNA only panels, the requirements are reduced to half
Specimen Type (select all that app	oly)					
Biopsy Type: FFPE Tissu	ue 🔲 F	F Tissue	Other	(specify)		
PARAFFIN BLOCK – no prep	ped scrolls	or extracte	d nucleic acids			
• DNA(ng)	RNA	(n	g)			
and may only be used for the deterr Nucleic must acid be extracted from All nucleic acids will be tested for qu FF and FFPE Tissue Recommendations	centrations and mination of same in a minimum wality as per law in-frozen in liquon	ample purity of 1 ml of bid aboratory the uid nitrogen a	(260/280 ≥ 1.8 for DNA a ppsy in EDTA, 120 μm or α resholds prior to processi as quickly as possible aftα	and ≥ 1.9 for of FFPE tissu ing er removal fr	RNA) le or 4 mm ³ of FF tissue	ediately delivered to the laboratory. Samples must be kept in -80° C
SPECIMEN TYPE		SHIP	PING & HANDLING INST	RUCTIONS		REJECTION CRITERIA
DNA & RNA	• Shir	• Ship at -20°C (use dry ice)		. 6	hand and a second for all a	
FF Tissue	·					optimal quantity/quality /FF: Tissue content < 40%; Tumor cellularity < 10%
FFPE Tissue	• Ship	at room te	emperature			
CHECKLIST						
A completed requisition has been sent with the specimen/s						
A pathology report has been sent with the specimen/s						
Any available genomic (single gene or panel) profile report/s has been sent with the specimen/s						
Please provide the following inform	Please provide the following information:					
Tissue content:			Tumor cellularity:			Pathologist's Name:

 ${\it FFPE: Formal in Fixed Paraffin Embedded \ tissue \ or \ block; FF \ Tissue: Fresh \ Frozen \ tissue}$

C3 Policy on Molecular Testing and Retesting Eligibility

Retesting will not be conducted on samples that have undergone prior molecular testing using a validated assay that is capable of detecting fusions, rearrangements, and covers the majority of mutations that are clinically actionable and relevant for biliary tract cancers. Additionally, retesting may not be conducted if a targetable mutation has already been identified, including but not limited to: KRAS, NRAS, BRAF, and IDH1/2 mutations. Furthermore, retesting may not be conducted if a patient is currently on a clinical trial with a targeted therapy for a mutation that was previously identified.

Please note, each test requisition will be evaluated on a case-by-case basis to determine eligibility for retesting. This assessment will include the criteria outlined above, as well as consideration of external factors such as the availability of funding.

The determination of test eligibility is at the sole discretion of the C3. Please be aware that these conditions may be subject to change or updates in the future.





SOLID TUMOR NGS PANEL DESCRIPTION

OncoHelix-2: 170 genes CGP Panel (DNA +/- RNA)

CGP Assay uses Illumina TST-170 panel*

Specimen compatibility: Genomic DNA & RNA extracted from fresh frozen and FFPE tissues

Small variants and indel (148): AKT1, AKT2, AKT3, ALK, APC, AR, ARID1A, ATM, ATR, BAP1, BARD1, BCL2, BCL6, BRAF, BRCA1, BRCA2, BRIP1, BTK, CARD11, CCND1, CCND2, CCNE1, CD79A, CD79B, CDH1, CDK12, CDK4, CDK6, CDKN2A, CEBPA, CHEK1, CHEK2, CREBBP, CSF1R, CTNNB1, DDR2, DNMT3A, EGFR, EP300, ERBB2, ERBB3, ERBB4, ERCC1, ERG, ESR1, EZH2, FAM175A, FANCI, FANCI, FBXW7, FGF1, FGF10, FGF2, FGF3, FGF3, FGF4, FGF5, FGF7, FGF9, FGFR1, FGFR3, FGFR4, FLT1, FLT3, FOXL2, GNA11, GNAQ, GNAS, HNF1A, HRAS, IDH1, IDH2, INPP4B, JAK2, JAK3, KDR, KIT, KRAS, MAP2K1, MAP2K2, MCL1, MDM2, MDM4, MET, MLH1, MLLT3, MPL, MRE11A, MSH2, MSH3, MSH6, MTOR, MUTYH, MYC, MYCN, MYD88, NBN, NF1, NOTCH1, NOTCH2, NOTCH3, NPM1, NRAS, NRG1, PALB2, PDGFRA, PDGFRB, PIK3CA, PIK3CB, PIK3CD, PIK3CG, PIK3R1, PMS2, PTCH1, PTEN, PTPN11, RAD51B, RAD51C, RAD54L, RB1, RET, RICTOR, ROS1, SLX4, SMAD4, SMARCB1, SMO, STK11, TET2, TP53, TSC1, TSC2. DNA amplification target genes (59): AKT2, ALK, AR, ATM, BRAF, BRCA1, BRCA2, CCND1, CCND3, CCNE1, CDK4, CDK6, CHEK1, CHEK2, EGFR, ERBB2, ERBB3, ERCC1, ERCC2, ESR1, FGF1, FGF14, FGF19, FGF2, FGF23, FGF3, FGF4, FGF5, FGF6, FGF7, FGF8, FGF9, FGFR1, FGFR2, FGFR3, FGFR4, JAK2, KIT, KRAS, LAMP1, MDM2, MDM4, MET, MYC, MYCL1, MYCN, NRAS, NRG1, PDGFRA, PDGFRB, PIK3CA, PIK3CB, PTEN, RAF1, RET, RICTOR, RPS6KB1, TFR. RNA fusion target genes (55): ABL1, AKT3, ALK, AR, AXL, BCL2, BRAF, BRCA1, BRCA2, CDK4, CSF1R, EGFR, EML4, ERBB2, ERG, ESR1, ETS1, ETV1, ETV4, ETV5, EWSR1, FGFR1, FGFR2, FGFR3, FGFR4, FLI1, FLT1, FLT3, JAK2, KDR, KIF5B, KIT, KMT2A (MLL), MET, MLLT3, MSH2, MYC, NOTCH1, NOTCH2, NOTCH3, NRG1, NTRK1, NTRK2, NTRK3, PAX3, PAX7, PDGFRA, PDGFRB, PIK3CA, PPARG, RAF1, RET, ROS1, RPS6KB1, TMPRSS2

*OncoHelix-2/3: 170 Gene CGP Panel uses the Illumina TST170 to provide comprehensive genomic profiling. The research use only assay was validated and its performance characteristics were determined by OncoHelix and its partner lab - Hematology Translational Lab. The panel is not approved by Health Canada, as is the case for all cancer genomic panel. Both OncoHelix and HTL laboratories are clinically accredited by CPSA to perform high-complexity molecular testing. Any decisions related to patient care and treatment choices should be based on the independent judgement of the treating physician

Testing Site & Shipping Address	For HTL Laboratory Use Only		
ATTN: Dr. Faisal Khan	Sample Received (YYYY-MM-DD) (AM/PM)		
Hematology Translational Lab (HTL)	Specimen type		
HMRB 336, 3330, Hospital Drive NW,	# Tubes/amount		
Calgary, AB, CANADA T2N 4N1	Lab Acc.#		